

RESEARCH TOPIC PRIME4

Mechanostimulation and antibiotic response in biofilm-forming bacteria under fluid flow

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Main facility

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Main field of interest

Biophysics; Microbiology

Abstract

Bacteria often organize into surface-attached communities known as biofilms, which readily form on implanted medical devices and are among the most common sources of hospital-acquired infections¹. Clinical isolates, such as those from biliary stents², display an enhanced capacity for biofilm formation, particularly under dynamic fluid conditions³. This points to an underexplored role of bacterial mechanosensitive channels, which are increasingly linked to infection dynamics and antibiotic resistance. To address this, we propose using recently developed membrane-targeting azobenzene photoswitches^{4,5} as remote, non-invasive optomechanical tools for the spatiotemporal control and interrogation of bacterial membrane potential, under fluid-induced mechanical stress in microfluidic systems. Complementarily, we aim to assess how such stress influences bacterial susceptibility to antibiotics. This multidisciplinary approach promises new insights into bacterial adaptation in clinically relevant settings and may guide the development of alternative strategies to combat antibiotic-resistant infections.

Main technical approaches

This project will combine advanced microscopy, microfluidic techniques, and quantitative image analysis to investigate bacterial responses to mechanical stress. Fluorescent optical microscopy and live-cell imaging will be employed to monitor bacterial membrane potential dynamics, biofilm development, and spatial organization in real-time. Custom-designed microfluidic devices will mimic physiologically relevant flow conditions, allowing precise control over mechanical stress applied to bacterial populations. These devices will also enable localized photoactivation of azobenzene-based photoswitches for targeted membrane stimulation. High-resolution image datasets acquired during experiments will be

analyzed using automated image processing and quantification pipelines to extract metrics such as biofilm density, morphology, membrane perturbation, and antibiotic susceptibility.

Scientific references

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Type of contract

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